C. DIFF QUIK CHEK COMPLETE®

A Rapid Membrane Enzyme Immunoassay for the Simultaneous Detection of Clostridium difficile Glutamate Dehydrogenase Antigen and Toxins A and B in Fecal Specimens

Catalog No. 30525C (25 Tests) or 30550C (50 Tests)

IVD In Vitro Diagnostic Medical Device

U.S. CLIA classification — Moderate

U. S. Patent #8,343,726

Developed and Manufactured by:

TECHLAB®, Inc.
2001 Kraft Drive
Blacksburg, VA 24060-6358
USA

Distributed by:

Alere North America, LLC
30 South Keller Road
Orlando, Florida 32810
TEL 1-877-441-7440
1-321-441-7200 OUTSIDE USA

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Made in the USA

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**INTENDED USE**

The C. DIFF QUIK CHEK COMPLETE™ test is a rapid membrane enzyme immunoassay for the simultaneous detection of Clostridium difficile glutamate dehydrogenase antigen and toxins A and B in a single reaction well. The test detects C. difficile antigen, glutamate dehydrogenase, as a screen for the presence of C. difficile and confirms the presence of toxigenic C. difficile by detecting toxins A and B in fecal specimens from persons suspected of having C. difficile disease. The test is to be used as an aid in the diagnosis of C. difficile disease. As with other C. difficile tests, results should be considered in conjunction with the patient history.

**EXPLANATION**

After treatment with antibiotics, many patients develop gastrointestinal problems ranging from mild diarrhea to severe pseudomembranous colitis. Many cases of the milder forms of gastrointestinal illness and most cases of pseudomembranous colitis are caused by toxigenic strains of Clostridium difficile (1). This organism is an opportunistic anaerobic bacterium that grows in the intestine once the normal flora has been altered by the antibiotic. Toxigenic strains of C. difficile carry the genes encoding the toxins while non-toxigenic strains do not carry the toxin genes. Disease onset is associated with the toxins that are produced by the toxigenic organism. The clinical symptoms associated with the disease are believed to be primarily due to toxin A, which is a tissue-damaging enterotoxin (2,3). C. difficile also produces a second toxin, designated toxin B. Toxin B, which has been referred to as the cytoxin of the organism, is the toxin detected by the tissue culture assay currently used by many laboratories. Toxigenic C. difficile strains produce both toxins, or only toxin B (4-7). The glutamate dehydrogenase of C. difficile is a good antigen marker for the organism in feces because it is produced in high amounts by all strains, toxigenic or non-toxigenic (8-10). The antigen can be detected in fecal specimens by using the C. DIFF QUIK CHEK COMPLETE™ test. A positive result in the test for the glutamate dehydrogenase of C. difficile confirms the presence of this organism in a fecal specimen; a negative result indicates the absence of the organism. A positive result in the test for toxins A and B confirms the presence of toxigenic C. difficile.

**PRINCIPLE OF THE TEST**

The C. DIFF QUIK CHEK COMPLETE™ test uses antibodies specific for glutamate dehydrogenase and toxins A and B of C. difficile. The device contains a Reaction Window with three vertical lines of immobilized antibodies. The antigen test line ("Ag") contains antibodies against C. difficile glutamate dehydrogenase. The control line ("C") is a dotted line that contains anti-horseradish peroxidase (HRP) antibodies. The toxins A and B test line ("Tox") contains antibodies against C. difficile toxins A and B. The Conjugate consists of antibodies to glutamate dehydrogenase and antibodies to toxins A and B coupled to horseradish peroxidase. To perform the test, the sample is added to a tube containing a mixture of Diluent and Conjugate. The diluted sample-conjugate mixture is added to the Sample Well and the device is allowed to incubate at room temperature for 15 minutes. During the incubation, any glutamate dehydrogenase and toxins A and B in the sample bind to the antibody-peroxidase conjugates. The antigen-antibody-conjugate complexes migrate through a filter pad to a membrane where they are captured by the immobilized glutamate dehydrogenase-specific and toxins A and B-specific antibodies in the lines. The Reaction Window is subsequently washed with Wash Buffer, followed by the addition of Substrate. After a 10 minute incubation period, the "Ag" reaction is examined visually for the appearance of a vertical blue line on the "Ag" side of the Reaction Window. A blue line indicates a positive test. If the "Ag" is positive, then the "Tox" reaction should be examined visually for the appearance of a blue line on the "Tox" side of the Reaction Window. A blue line indicates a positive test. A positive "C" reaction, indicated by a vertical dotted blue line under the "C" portion of the Reaction Window, confirms that the test is working properly and the results are valid.
REFERENCES


MATERIALS PROVIDED

<table>
<thead>
<tr>
<th>Material</th>
</tr>
</thead>
<tbody>
<tr>
<td>Membrane Devices – each pouch contains 1 device</td>
</tr>
<tr>
<td>Diluent (22 mL per bottle) – Buffeted protein solution with graduated dropper assembly (contains 0.05% ProClin® 300)</td>
</tr>
<tr>
<td>Signal Word: Warning</td>
</tr>
<tr>
<td>H317: May cause an allergic skin reaction</td>
</tr>
<tr>
<td>P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501</td>
</tr>
<tr>
<td>Wash Buffer (12 mL per bottle) – Buffeted solution with graduated dropper assembly (contains 0.05% ProClin® 300)</td>
</tr>
<tr>
<td>Signal Word: Warning</td>
</tr>
<tr>
<td>H317: May cause an allergic skin reaction</td>
</tr>
<tr>
<td>P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501</td>
</tr>
<tr>
<td>Substrate (3.5 mL per bottle) – Solution containing tetramethylbenzidine Conjugate (2.5 mL per bottle) – Mouse monoclonal antibody specific for glutamate dehydrogenase coupled to horseradish peroxidase and goat polyclonal antibodies specific for toxins A and B coupled to horseradish peroxidase in a buffered protein solution</td>
</tr>
<tr>
<td>Contains 0.05% ProClin® 300</td>
</tr>
<tr>
<td>Signal Word: Warning</td>
</tr>
<tr>
<td>H317: May cause an allergic skin reaction</td>
</tr>
<tr>
<td>P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501</td>
</tr>
<tr>
<td>Positive Control (2 mL) – Antigen in a buffered protein solution</td>
</tr>
</tbody>
</table>

Disposable plastic transfer pipettes – graduated at 25 µL, 400 µL and 500 µL

In Vitro Diagnostic Medical Device

MATERIALS AND EQUIPMENT REQUIRED BUT NOT PROVIDED

- Small test tubes (e.g., plastic Eppendorf tubes or glass tubes)
- Applicator sticks
- Vortex mixer
- Disposable gloves for handling fecal samples
- Pipettor and tips

SHELF LIFE AND STORAGE

The expiration date of the kit is given on the label. Expiration dates for each component are listed on the individual labels. The kit should be stored between 2°C and 8°C.

PRECAUTIONS

1. Reagents from different kits should not be mixed or interchanged. Do not use a kit past the expiration date.
2. Each component in the kit should be inspected for any signs of leakage. Upon arrival, inspect the kit to ensure that components are not frozen or warm to the touch due to improper shipping conditions.
3. Bring all components to ROOM TEMPERATURE BEFORE USE!
4. Caps, tips and dropper assemblies are color-coded; do NOT mix or interchange!
5. Do not freeze the reagents. The kit should be stored between 2°C and 8°C.
6. The pouch containing the Membrane Device should be at room temperature before opening. Keep the membrane devices dry before use.
7. Use fecal specimens within 72 hours of collection to obtain optimal results. Specimens that are frozen may lose activity due to freezing and thawing. If using frozen specimens, thaw at room temperature.
8. Hold reagent bottles vertically to dispense reagents to ensure consistent drop size and correct volume.
9. Specimens and membrane devices should be handled and disposed of as potential biohazards after use.
10. Membrane devices cannot be reused.
The test has been optimized for sensitivity and specificity. Alterations of the specified procedure and/or test conditions may affect the sensitivity and specificity of the test. Do not deviate from the specified procedure.

11. Be attentive to the total assay time when testing more than one fecal specimen. Add Diluent first, and then add the Conjugate to each tube of Diluent. Then add specimen to the tube of Diluent/Conjugate. Thoroughly mix all of the diluted specimens, and transfer to the Membrane Device. The 15-minute incubation step begins after the last diluted sample-conjugate mixture has been transferred to the final Membrane Device.

12. If the Substrate reagent changes to a dark blue/violet color call technical services for replacement.

13. Fecal specimens may contain potentially infectious agents and should be handled at “Biosafety Level 2” as recommended in the CDC/NIH Manual “Biosafety in Microbiological and Biomedical Laboratories.”

14. All reagents are for in vitro diagnostic use only.

15. Wear disposable gloves when doing the test.

16. The following substances (U.S. formulation) had no effect on test results when present in feces in the concentrations indicated: mucin (3.5% w/v), human blood (40% v/v), barium sulfate (5% w/v), Imodium® (5% v/v), Kaopectate® (5% v/v), Pepto-Bismol® (5% v/v), steric/palmitic acid (40% w/v), Metronidazole (0.25% w/v), Vancomycin (0.25% w/v).

17. Reaction of Clinical Isolates Obtained on Cycloserine-Cefoxitin-Fructose Agar (CCFA)

A total of 103 C. difficile clinical isolates, obtained by anaerobic bacterial culture on CCFA after 3 days at 37°C, were tested in the C. DIFF QUIK CHEK COMPLETE® test. For the analysis, individual colonies were picked and suspended in Diluent as recommended for fecal specimens. All 103 isolates gave a positive antigen reaction in the test. Seventy of the 103 isolates (68%) were from fecal specimens that were positive for C. difficile toxin by tissue culture assay. Of these, 56 (80%) gave a positive toxin reaction when screened following anaerobic growth on CCFA for 3 days at 37°C.

The only non-C. difficile organism to react in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® test was Clostridium sordellii VPI 9048. This strain produces toxins HT and LT, which are homologous to toxins A and B, respectively.

The following viruses of 10^2.1 to 10^2.25 TCID units per 0.2 mL did not react in the C. DIFF QUIK CHEK COMPLETE® test:

- Adenovirus types 1, 2, 3, 5, 40, 41
- Human coronavirus
- Coxsackievirus B2, B3, B4, B5
- Echovirus 9, 11, 18, 22, 33
- Enterovirus type 68, 69, 70, 71
- Rotavirus

INTERFERING SUBSTANCES

The following substances (U.S. formulation) had no effect on test results when present in feces in the concentrations indicated: mucin (3.5% w/v), human blood (40% v/v), barium sulfate (5% w/v), Imodium® (5% v/v), Kaopectate® (5% v/v), Pepto-Bismol® (5% v/v), steric/palmitic acid (40% w/v), Metronidazole (0.25% w/v), Vancomycin (0.25% w/v).

COLLECTION, HANDLING, AND STORAGE OF FECAL SPECIMENS

<table>
<thead>
<tr>
<th>Acceptable Sample Types</th>
<th>Do Not Use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fresh Fecal Specimens</td>
<td>Fecal specimens in Formalin-based fixative (e.g. sodium acetate formalin, 10% formalin, merthiolate formalin)</td>
</tr>
<tr>
<td>Frozen Fecal Specimens</td>
<td>Fecal specimens in alcohol-based fixative (e.g. polyvinyl alcohol)</td>
</tr>
<tr>
<td>Specimens in transport media (Cary Blair, C&amp;S)</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Sample Storage Temperature</th>
<th>Acceptable length of storage</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>2°C – 8°C</td>
<td>72 hours</td>
<td>Ideal specimens are less than 24 hours old</td>
</tr>
<tr>
<td>Frozen ≤ -10°C</td>
<td>Longer than 72 hours</td>
<td>Thaw at room temperature. Freezing and thawing multiple times may result in loss of specimen activity due to toxin degradation.</td>
</tr>
</tbody>
</table>

1. Standard collection and handling procedures used in-house for fecal specimens are appropriate.
2. Fecal specimens should be collected in clean, leak-proof containers.
3. Storing fecal specimens in the Diluent is NOT recommended.
4. Do not allow the fecal specimens to remain in the Diluent/Conjugate mixture for >24 hours.
ANALYTICAL SENSITIVITY
The cutoff for the assay was established at concentrations of 0.63 ng/mL for toxin A, 0.16 ng/mL for toxin B, and 0.8 ng/mL for glutamate dehydrogenase.

REPRODUCIBILITY
The reproducibility of the C. DIFF QUIK CHEK COMPLETE® test was determined using 12 fecal specimens that were coded to prevent their identification during testing. Testing was performed at 3 independent laboratories, which tested the samples for 3 days. The samples produced the expected results 100% of the time.

An additional 5-day study was performed at 3 sites by running a set of low positive, moderate positive and high negative fecal samples that were spiked with glutamate dehydrogenase, Toxin A and Toxin B. The samples were run in triplicate, twice a day over a 5-day period by multiple technicians at each site. The combined antigen and toxin data from the 5-day reproducibility study is shown in Table 6. The antigen data for Sample 1 was below 90% negative at one site and was consistently negative for antigen at the other two sites. The toxin data for Sample A was below 90% positive at one site, and was consistently positive for toxin at the other two sites. No single site reported results below expectations for both antigen and toxin.

Table 6. Summary of 5-day reproducibility study.

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Antigen Positive</th>
<th>Antigen Negative</th>
<th>Toxin Positive</th>
<th>Toxin Negative</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample 1 (95% negative results expected)</td>
<td>15 (16.7%)</td>
<td>75 (83.3%)</td>
<td>0 (0%)</td>
<td>90 (100%)</td>
</tr>
<tr>
<td>Sample A (95% positive results expected)</td>
<td>89 (98.9%)</td>
<td>1 (1.1%)</td>
<td>65 (72.2%)</td>
<td>25 (27.8%)</td>
</tr>
<tr>
<td>Sample B (mod positive)</td>
<td>87 (96.7%)</td>
<td>3 (3.3%)</td>
<td>86 (95.5%)</td>
<td>4 (4.5%)</td>
</tr>
</tbody>
</table>

CROSS REACTIVITY
Fecal specimens inoculated with the following microorganisms to a final concentration of approximately 10^8 or higher organisms per mL did not react in the antigen or toxin portion of the C. DIFF QUIK CHEK COMPLETE® test:

**Bacterium or Pathogen:** Aeromonas hydrophila, Bacillus cereus, Bacillus subtilis, Bacteroides fragilis, Campylobacter coli, Campylobacter fetus, Campylobacter jejuni, Candida albicans, Clostridium butyricum, Clostridium clostridiforme, Clostridium haemolyticum, Clostridium histolyticum, Clostridium novyi, Clostridium perfringens, Clostridium septicum, Clostridium sordellii (nontoxigenic), Clostridium sporogenes, Enterobacter aerogenes, Enterobacter cloacae, Enterococcus faecalis, Escherichia coli EIEC, Escherichia coli, Escherichia coli O157:H7, Escherichia coli ETEC, Klebsiella pneumoniae, Pseudostreptococcus anaerobius, Proteus vulgaris, Pseudomonas aeruginosa, Salmonella typhimurium, Seratia liquefaciens, Shigella dysenteriae, Shigella flexneri, Shigella sonnei, Staphylococcus aureus, Staphylococcus aureus (Cowans), Staphylococcus epidermidis, Vibrio cholerae, Vibrio parahaemolyticus, Yersinia enterocolitica

SPECIMEN PREPARATION
1. Bring all reagents and the required number of devices to room temperature before use. It is recommended to remove the reagents from the foam insert to reduce the time needed to warm to room temperature.
2. Set up and label one small test tube for each specimen, and optional external controls as necessary.
3. Using the black graduated dropper assembly, add 750 µL (2nd graduation from the tip) Diluent to each tube for fecal specimens. For specimens in transport media such as Cary Blair or C&S, add 650 µL (1st graduation from the tip) of Diluent to the tube.
4. Add one drop of Conjugate (red capped bottle) to each tube.
5. Obtain one disposable plastic transfer pipette (supplied with the kit) for each sample – the pipettes have raised graduations at 25 µL, 400 µL and 500 µL.

Graduated Transfer Pipette:

6. Mix all specimens thoroughly regardless of consistency - it is essential that the specimens be evenly suspended before transferring. Liquid/Semi-solid specimens – pipette 25 µL of specimen with a transfer pipette and dispense into the Diluent/Conjugate mixture. Use the same transfer pipette to mix the diluted specimen.

Formed/Solid specimens – Care must be taken to add the correct amount of formed feces to the sample mixture. Mix the specimen thoroughly using a wooden applicator stick and transfer a small portion (approximately 2 mm diameter, the equivalent of 25 µL) of the specimen into the Diluent/Conjugate mixture. Emulsify the specimen using the applicator stick.

Fecal specimens in Cary Blair or C&S transport media - pipette 100 µL (2 drops from transfer pipette) of sample into the Diluent/Conjugate mixture.

7. Optional External Control Samples:

**External Positive Control** - add one drop of Positive Control (gray-capped bottle) into the Diluent/Conjugate mixture.

**External Negative Control** - add 25 µL Diluent into the Diluent/Conjugate mixture.

NOTE: Transferring too little specimen, or failure to mix and completely suspend the specimen in the Diluent/Conjugate mixture, may result in a false-negative test result. The addition of too much fecal specimen may cause invalid results due to restricted sample flow.
TEST PROCEDURE
1. Obtain one Membrane Device per specimen, and one device per optional external positive or negative control as necessary. The foil bags containing the devices should be brought to room temperature before opening. Use the device immediately after opening. Label each device appropriately and orient it on a flat surface so the “C. DIFF COMPLETE” print is at the bottom of the device, and the small Sample Well is located in the top right corner of the device.

2. Close each tube of diluted specimen and mix thoroughly. Proper mixing can be achieved by vortexing or inverting the tube. Once a patient sample or Positive Control has been diluted in the Diluent/Conjugate mixture, it may be incubated at room temperature for any period of time up to 24 hours prior to addition to the Membrane Device.

3. Using a new transfer pipette, transfer 500 µL of the diluted sample-conjugate mixture into the Sample Well (smaller hole in the top right corner of the device) of a Membrane Device, making certain to expel the liquid sample onto the wicking pad inside of the Membrane Device. When loading the sample into the sample well, make sure that the tip of the transfer pipette is angled towards the Reaction Window (larger hole in the middle of the device).

4. Incubate the device at room temperature for 15 minutes – the sample will wick through the device and a wet area will spread across the Reaction Window.

NOTE FOR SAMPLES THAT FAIL TO MIGRATE:
Occasionally, a diluted sample fails to migrate properly and the Reaction Window does not fully wet. If the Reaction Window does not appear to be completely wet within 5 minutes of adding the sample to the Sample Well, then add 100 µL (4 drops) of Diluent to the Sample Well and wait an additional 5 minutes (for a total of 20 minutes).

5. After the incubation, add 300 µL of Wash Buffer to the Reaction Window using the graduated white dropper assembly. Allow the Wash Buffer to flow through the Reaction Window membrane and be absorbed completely.

6. Add 2 drops of Substrate (white-capped bottle) to the Reaction Window. Read and record results visually after 10 minutes.

INTERPRETATION OF RESULTS
1. Interpretation of the test is most reliable when the device is read immediately at the end of the 10 minute reaction period. Read the device at a normal working distance in a well-lit area. View with a line of vision directly over the device.

2. Observe device for the appearance of blue dots in the middle of the Reaction Window representing the internal positive control. The appearance of any control dot(s) represents a valid internal control. The background may appear white to light blue in color. Observe device for the appearance of blue lines on the “Ag” and “Tox” sides of the Reaction Window representing the test lines. The lines may appear faint to dark in intensity.

EFFECT OF FECAL SPECIMEN CONSISTENCY
Effect of fecal specimen consistency on the C. DIFF QUIK CHEK COMPLETE® test
The reaction of fecal specimens of varying consistencies in the antigen portion (n=978) and toxin portion (n=981) of the C. DIFF QUIK CHEK COMPLETE® test is shown in Tables 4 and 5. The percentages of positive reactions using either culture assay or the C. DIFF QUIK CHEK COMPLETE® test were similar in all three types of fecal specimens (liquid, semi-solid, and solid). All of the specimens were submitted for C. difficile testing. The basis of the submission was the clinical history of the patient and not the consistency of the specimen. In the antigen portion, the results show that the C. DIFF QUIK CHEK COMPLETE® test performed similarly to bacterial culture when testing samples of different consistencies. In the toxin portion, the results show the C. DIFF QUIK CHEK COMPLETE® test performed similarly to the tissue culture assay when testing samples of different consistencies.

Table 4. Reaction of fecal specimens of varying consistencies in the antigen portion of the C. DIFF QUIK CHEK COMPLETE® test

<table>
<thead>
<tr>
<th>Type of Specimen</th>
<th>Liquid Specimens (n = 978)</th>
<th>Semi-solid Specimens (n = 522)</th>
<th>Solid Specimens (n = 121)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive by bacterial culture assay</td>
<td>59 (17.6%)</td>
<td>110 (21.1%)</td>
<td>19 (15.7%)</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Positive</td>
<td>72 (21.5%)</td>
<td>128 (24.5%)</td>
<td>25 (20.7%)</td>
</tr>
<tr>
<td>Negative by bacterial culture assay</td>
<td>276 (82.4%)</td>
<td>412 (78.9%)</td>
<td>102 (84.3%)</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Negative</td>
<td>263 (78.5%)</td>
<td>394 (75.5%)</td>
<td>96 (79.3%)</td>
</tr>
</tbody>
</table>

Table 5. Reaction of fecal specimens of varying consistencies in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® test

<table>
<thead>
<tr>
<th>Type of Specimen</th>
<th>Liquid Specimens (n = 981)</th>
<th>Semi-solid Specimens (n = 523)</th>
<th>Solid Specimens (n = 122)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive by tissue culture assay</td>
<td>43 (12.8%)</td>
<td>81 (15.5%)</td>
<td>8 (6.6%)</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Toxin Line Positive</td>
<td>42 (12.5%)</td>
<td>72 (13.8%)</td>
<td>7 (5.7%)</td>
</tr>
<tr>
<td>Negative by tissue culture assay</td>
<td>293 (87.2%)</td>
<td>442 (84.5%)</td>
<td>114 (93.4%)</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Toxin Line Negative</td>
<td>294 (87.5%)</td>
<td>451 (86.2%)</td>
<td>115 (94.3%)</td>
</tr>
</tbody>
</table>
The antigen portion of the *C. DIFF QUIK CHEK COMPLETE*® test was compared to the tissue culture assay for the detection of *C. difficile* toxin. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The results are shown in Table 2. The antigen portion of the *C. DIFF QUIK CHEK COMPLETE*® test detected 98.7% of the tissue culture-positive samples.

Table 2. Summary of clinical performance comparing *C. DIFF QUIK CHEK COMPLETE*® test to the tissue culture assay

<table>
<thead>
<tr>
<th></th>
<th>Tissue Culture positive</th>
<th>Tissue Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>C. DIFF QUIK CHEK COMPLETE®</strong> Antigen Line Positive</td>
<td>154</td>
<td>109</td>
</tr>
<tr>
<td><strong>C. DIFF QUIK CHEK COMPLETE®</strong> Antigen Line Negative</td>
<td>2</td>
<td>861</td>
</tr>
</tbody>
</table>

Clinical evaluation of the toxin portion of the *C. DIFF QUIK CHEK COMPLETE*® test

The toxin portion of the *C. DIFF QUIK CHEK COMPLETE*® test was compared to the tissue culture assay at two clinical laboratories and in-house at TECHLAB®, Inc. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The results are shown in Table 3.

Table 3. Summary of clinical performance comparing *C. DIFF QUIK CHEK COMPLETE*® test to the tissue culture assay

<table>
<thead>
<tr>
<th></th>
<th>Tissue Culture positive</th>
<th>Tissue Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>C. DIFF QUIK CHEK COMPLETE®</strong> Toxin Line Positive</td>
<td>137</td>
<td>6</td>
</tr>
<tr>
<td><strong>C. DIFF QUIK CHEK COMPLETE®</strong> Toxin Line Negative</td>
<td>19</td>
<td>964</td>
</tr>
</tbody>
</table>

95% Confidence Limits

- Sensitivity: 87.8% (81.4 - 92.3)
- Specificity: 99.4% (98.6 - 99.7)
- Predictive Positive Value: 95.8% (90.7 - 98.3)
- Predictive Negative Value: 98.1% (96.9 - 98.8)
- Correlation: 97.8% (97.6 - 98.0)

Discrepant samples were evaluated using current ELISA tests for toxins A and B. Five of the 6 false positive samples were positive by ELISA and were considered true positives. Twelve of the 19 false negative samples were negative by ELISA and were considered true negatives.

3. **Positive Antigen (“Ag”) Result:** A positive antigen result may be interpreted at any time between the addition of Substrate and the 10-minute read time. For a positive antigen result, the blue “Ag” line and the dotted blue control line below “C” are visible (Figure 1a). The lines may appear faint to dark in intensity. An obvious partial line is interpreted as a positive result. Do not interpret membrane discoloration as a positive result. A positive result indicates the presence of *C. difficile*.

4. **Positive Antigen and Toxin (“Tox”) Result:** If the antigen result is positive (i.e., a blue “Ag” line and a dotted blue control below “C” are visible), proceed to the interpretation of the toxin result. A positive toxin result may be interpreted at any time between the addition of Substrate and the 10-minute read time. For a positive toxin result, a blue “Tox” line is visible (Figure 1b). The line may appear faint to dark in intensity. An obvious partial line is interpreted as a positive result. Do not interpret membrane discoloration as a positive result. A positive result indicates the presence of *C. difficile* toxin.

5. **Negative Result:** A test cannot be interpreted as negative or invalid until 10 minutes following the addition of Substrate. A single blue dotted line is visible in the middle of the Reaction Window, below the “C” and no test lines are visible on the “Ag” side or the “Tox” side of the Reaction Window (Figure 1c). A negative result in the antigen portion indicates *C. difficile* antigen is either absent in the specimen or is below the detection limit of the test. A negative result in the toxin portion indicates *C. difficile* toxin is either absent in the specimen or is below the detection limit of the test.

6. **Invalid Result:** No lines are visible in the Reaction Window (Figure 1d). The test result is invalid if a blue dotted line is not present below the “C” at the completion of the reaction period (Figures 1e, 1f, 1g).

7. **Negative Antigen (“Ag”), Positive Toxin (“Tox”):** A low percentage of specimens may test negative for antigen but positive for toxin. These samples should be considered indeterminate and retested using a fresh specimen (Figure 1h). If sample retests negative for antigen but positive for toxin, report as positive toxin result.
QUALITY CONTROL

Internal: A dotted blue line must be visible in the middle of the Reaction Window, below the “C” on every Membrane Device that is tested. The appearance of the blue control dots confirms that the sample and reagents were added correctly, that the reagents were active at the time of performing the assay, and that the sample migrated properly through the Membrane Device. It also confirms the reactivity of the other reagents associated with the assay. A clear background in the result area is considered an internal negative control. If the test has been performed correctly and reagents are working properly, the background will be white to give a discernible result.

External: The reactivity of the C. DIFF QUIK CHEK COMPLETE® kit should be verified upon receipt using the Positive Control and negative control (Diluent). The Positive Control is supplied with the kit (gray-capped bottle). The Positive Control confirms the reactivity of the other reagents associated with the assay, and is not intended to ensure precision at the analytical assay cut-off. Diluent is used for the negative control. Additional tests can be performed with the controls to meet the requirements of local, state and/or federal regulations and/or accrediting organizations.

LIMITATIONS

1. The C. DIFF QUIK CHEK COMPLETE® test is used to detect C. difficile antigen and toxin(s) in fecal specimens. The test confirms the presence of toxin in feces and this information should be taken under consideration by the physician in light of the clinical history and physical examination of the patient. The C. DIFF QUIK CHEK COMPLETE® test will detect levels of toxin A at ≥0.63 ng/mL, toxin B at ≥0.16 ng/mL, and glutamate dehydrogenase at ≥0.8 ng/mL.
2. Fecal specimens are extremely complex. Optimal results with the C. DIFF QUIK CHEK COMPLETE® test are obtained with specimens that are less than 24 hours old. Most undiluted specimens can be stored between 2°C and 8°C for 72 hours before significant degradation of the toxin is noted. If specimens are not assayed within this time period, they may be frozen and thawed. However, repeated freezing and thawing may result in loss of the immune reactivity of antigen and toxins A and B.
3. Some specimens may give weak reactions. This may be due to a number of factors such as: the presence of low levels of antigen and/or toxin, the presence of binding substances, or inactivating enzymes in the feces. The lines may appear faint to dark in intensity. These specimens should be reported as positive if any blue line, even a partial line is observed. An obvious partial blue line is interpreted as a positive result.
4. Fecal specimens preserved in 10% Formalin, merthiolate formalin, sodium acetate formalin, or polyvinyl alcohol cannot be used.
5. The C. DIFF QUIK CHEK COMPLETE® test is qualitative. The intensity of the color should not be interpreted quantitatively.
6. Some isolates of C. sordelli may react in the C. DIFF QUIK CHEK COMPLETE® test due to the production of immunologically related toxins (1).
7. Colonization rates of up to 50% have been reported in infants. A high rate has also been reported in cystic fibrosis patients (1,3). Results may appear positive in these groups but should be viewed in conjunction with the potential to be a colonized carrier.
8. The only non-C. difficile organism to react in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® test was Clostridium sordelli VPI 9048. This strain produces toxins HT and LT, which are homologous to toxins A and B, respectively.
9. No data exists on the effects of colonic washes, barium enemas, laxatives, or bowel preparations on the performance of the C. DIFF QUIK CHEK COMPLETE® test. All of these procedures can result in extensive dilution or the presence of additives that may affect test performance.

EXPECTED VALUES

Clostridium difficile disease is primarily a nosocomial disease of elderly patients, and the frequency of the disease is dependent on factors such as patient population, type of institution and epidemiology. The reported incidence of C. difficile disease in patients with antibiotic-associated diarrhea may range from 5 to 20%, and hospitals may experience rates higher or lower than this range. It is important to consider any test results in conjunction with clinical symptoms because some healthy adults and large numbers of healthy infants (up to 50%) will be positive for C. difficile toxin. In addition, C. difficile carriage rates of 22% to 32% have been reported in cystic fibrosis patients (1,3). In the studies conducted for this device, using symptomatic patients, the incidence of toxins A and B was 12% and GDH was 18%. A positive result in the antigen portion of the C. DIFF QUIK CHEK COMPLETE® test confirms the presence of C. difficile in a fecal specimen; a negative result indicates the absence of the organism. A positive result in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® confirms the presence of C. difficile toxin in a fecal specimen; a negative result indicates the absence of toxin or insufficient levels of toxin for detection.

PERFORMANCE CHARACTERISTICS

Clinical evaluation of the antigen portion of the C. DIFF QUIK CHEK COMPLETE® test

The antigen portion of the C. DIFF QUIK CHEK COMPLETE® test was compared to bacterial culture. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The bacterial culture test was performed according to in-house procedures. The results are shown in Table 1.

Table 1. Summary of clinical performance comparing C. DIFF QUIK CHEK COMPLETE® test to bacterial culture

<table>
<thead>
<tr>
<th>n = 1126</th>
<th>Bacterial Culture positive</th>
<th>Bacterial Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE®</td>
<td>201</td>
<td>62</td>
</tr>
<tr>
<td>Antigen Line Positive</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE®</td>
<td>21</td>
<td>842</td>
</tr>
<tr>
<td>Antigen Line Negative</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Sensitivity: 90.5% (95% CI: 85.7 - 93.9)
Specificity: 93.1% (95% CI: 91.2 - 94.7)
Predictive Positive Value: 76.4% (95% CI: 70.7 - 81.3)
Predictive Negative Value: 97.6% (95% CI: 96.2 - 98.4)
Correlation: 92.6% (95% CI: 91.8 - 93.4)

Discrepant samples were evaluated using current ELISA tests for C. difficile glutamate dehydrogenase. Twenty-nine of the 62 false positive samples were positive by another GDH test, and were considered true positives. Thirteen of the 21 false negative samples were negative by another GDH test, and were considered true negatives.
QUALITY CONTROL

Internal: A dotted blue line must be visible in the middle of the Reaction Window, below the “C” on every Membrane Device that is tested. The appearance of the blue control dots confirms that the sample and reagents were added correctly, that the reagents were active at the time of performing the assay, and that the sample migrated properly through the Membrane Device. It also confirms the reactivity of the other reagents associated with the assay. A clear background in the result area is considered an internal negative control. If the test has been performed correctly and reagents are working properly, the background will be white to give a discernible result.

External: The reactivity of the C. DIFF QUIK CHEK COMPLETE® kit should be verified upon receipt using the Positive Control and negative control (Diluent). The Positive Control is supplied with the kit (gray-capped bottle). The Positive Control confirms the reactivity of the other reagents associated with the assay, and is not intended to ensure precision at the analytical assay cut-off. Diluent is used for the negative control. Additional tests can be performed with the controls to meet the requirements of local, state and/or federal regulations and/or accrediting organizations.

LIMITATIONS
1. The C. DIFF QUIK CHEK COMPLETE® test is used to detect C. difficile antigen and toxin(s) in fecal specimens. The test confirms the presence of toxin in feces and this information should be taken under consideration by the physician in light of the clinical history and physical examination of the patient. The C. DIFF QUIK CHEK COMPLETE® test will detect levels of toxin A at ≥0.63 ng/mL, toxin B at ≥0.16 ng/mL, and glutamate dehydrogenase at ≥0.8 ng/mL.
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3. Some specimens may give weak reactions. This may be due to a number of factors such as the presence of low levels of antigen and/or toxin, the presence of binding substances, or inactivating enzymes in the feces. The lines may appear faint to dark in intensity. These specimens should be reported as positive if any blue line, even a partial line is observed. An obvious partial blue line is interpreted as a positive result.
4. Fecal specimens preserved in 10% Formalin, merthiolate formalin, sodium acetate formalin, or polyvinyl alcohol cannot be used.
5. The C. DIFF QUIK CHEK COMPLETE® test is qualitative. The intensity of the color should not be interpreted quantitatively.
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8. The only non-C. difficile organism to react in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® test was Clostridium sordelli VPI 9048. This strain produces toxins HT and LT, which are homologous to toxins A and B, respectively.
9. No data exists on the effects of colonic washes, barium enemas, laxatives, or bowel preparations on the performance of the C. DIFF QUIK CHEK COMPLETE® test. All of these procedures can result in extensive dilution or the presence of additives that may affect test performance.

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PERFORMANCE CHARACTERISTICS
Clinical evaluation of the antigen portion of the C. DIFF QUIK CHEK COMPLETE® test
The antigen portion of the C. DIFF QUIK CHEK COMPLETE® test was compared to bacterial culture. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The bacterial culture test was performed according to in-house procedures. The results are shown in Table 1.

Table 1. Summary of clinical performance comparing C. DIFF QUIK CHEK COMPLETE® test to bacterial culture

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<tr>
<th></th>
<th>n = 1126</th>
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<th>Bacterial Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Positive</td>
<td>201</td>
<td>62</td>
<td></td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Negative</td>
<td>21</td>
<td>842</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>Predictive Positive Value</th>
<th>Predictive Negative Value</th>
<th>Correlation</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>90.5%</td>
<td>93.1%</td>
<td>76.4%</td>
<td>97.6%</td>
<td>92.6%</td>
</tr>
</tbody>
</table>

Discrepant samples were evaluated using current ELISA tests for C. difficile glutamate dehydrogenase.

Twenty-nine of the 62 false positive samples were positive by another GDH test, and were considered true positives.

Thirteen of the 21 false negative samples were negative by another GDH test, and were considered true negatives.
The antigen portion of the *C. DIFF QUIK CHEK COMPLETE®* test was compared to the tissue culture assay for the detection of *C. difficile* toxin. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The results are shown in Table 2. The antigen portion of the *C. DIFF QUIK CHEK COMPLETE®* test detected 98.7% of the tissue culture-positive samples.

Table 2. Summary of clinical performance comparing *C. DIFF QUIK CHEK COMPLETE®* test to the tissue culture assay

<table>
<thead>
<tr>
<th></th>
<th>Tissue Culture positive</th>
<th>Tissue Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>C. DIFF QUIK CHEK COMPLETE®</em> Antigen Line Positive</td>
<td>154</td>
<td>109</td>
</tr>
<tr>
<td><em>C. DIFF QUIK CHEK COMPLETE®</em> Antigen Line Negative</td>
<td>2</td>
<td>861</td>
</tr>
</tbody>
</table>

Clinical evaluation of the toxin portion of the *C. DIFF QUIK CHEK COMPLETE®* test

The toxin portion of the *C. DIFF QUIK CHEK COMPLETE®* test was compared to the tissue culture assay at two clinical laboratories and in-house at TECHLAB®, Inc. Specimens included in the evaluation were submitted to the clinical laboratories for routine testing. The results are shown in Table 3.

Table 3. Summary of clinical performance comparing *C. DIFF QUIK CHEK COMPLETE®* test to the tissue culture assay

<table>
<thead>
<tr>
<th></th>
<th>Tissue Culture positive</th>
<th>Tissue Culture negative</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>C. DIFF QUIK CHEK COMPLETE®</em> Toxin Line Positive</td>
<td>137</td>
<td>6</td>
</tr>
<tr>
<td><em>C. DIFF QUIK CHEK COMPLETE®</em> Toxin Line Negative</td>
<td>19</td>
<td>964</td>
</tr>
</tbody>
</table>

95% Confidence Limits

- Sensitivity: 87.8% (81.4 - 92.3)
- Specificity: 99.4% (98.6 - 99.7)
- Predictive Positive Value: 95.8% (90.7 - 98.3)
- Predictive Negative Value: 98.1% (96.9 - 98.8)
- Correlation: 97.8% (97.6 - 98.0)

Discrepant samples were evaluated using current ELISA tests for toxins A and B. Five of the 6 false positive samples were positive by ELISA and were considered true positives. Twelve of the 19 false negative samples were negative by ELISA and were considered true negatives.

3. **Positive Antigen (“Ag”) Result:** A positive antigen result may be interpreted at any time between the addition of Substrate and the 10-minute read time. For a positive antigen result, the blue “Ag” line and the dotted blue control line below “C” are visible (Figure 1a). The lines may appear faint to dark in intensity. An obvious partial line is interpreted as a positive result. Do not interpret membrane discoloration as a positive result. A positive result indicates the presence of *C. difficile*.

4. **Positive Antigen and Toxin (“Tox”) Result:** If the antigen result is positive (i.e., a blue “Ag” line and a dotted blue control below “C” are visible), proceed to the interpretation of the toxin result. A positive toxin result may be interpreted at any time between the addition of Substrate and the 10-minute read time. For a positive toxin result, a blue “Tox” line is visible (Figure 1b). The line may appear faint to dark in intensity. An obvious partial line is interpreted as a positive result. Do not interpret membrane discoloration as a positive result. A positive result indicates the presence of *C. difficile* toxin.

5. **Negative Result:** A test cannot be interpreted as negative or invalid until 10 minutes following the addition of Substrate. A single blue dotted line is visible in the middle of the Reaction Window, below the “C” and no test lines are visible on the “Ag” side or the “Tox” side of the Reaction Window (Figure 1c). A negative result in the antigen portion indicates *C. difficile* antigen is either absent in the specimen or is below the detection limit of the test. A negative result in the toxin portion indicates *C. difficile* toxin is either absent in the specimen or is below the detection limit of the test.

6. **Invalid Result:** No lines are visible in the Reaction Window (Figure 1d). The test result is invalid if a blue dotted line is not present below the “C” at the completion of the reaction period (Figures 1e, 1f, 1g).

7. **Negative Antigen (“Ag”), Positive Toxin (“Tox”):** A low percentage of specimens may test negative for antigen but positive for toxin. These samples should be considered indeterminate and retested using a fresh specimen (Figure 1h). If sample retests negative for antigen but positive for toxin, report as positive toxin result.

**FIGURE 1:** *C. DIFF QUIK CHEK COMPLETE®* INTERPRETATION OF RESULTS

[Images of test results are shown here, including figures 1a to 1d, 1e to 1h.]
TEST PROCEDURE
1. Obtain one Membrane Device per specimen, and one device per optional external positive or negative control as necessary. The foil bags containing the devices should be brought to room temperature before opening. Use the device immediately after opening. Label each device appropriately and orient it on a flat surface so the “C. DIFF COMPLETE” print is at the bottom of the device, and the small Sample Well is located in the top right corner of the device.

![Membrane Device Diagram]

2. Close each tube of diluted specimen and mix thoroughly. Proper mixing can be achieved by vortexing or inverting the tube. Once a patient sample or Positive Control has been diluted in the Diluent/Conjugate mixture, it may be incubated at room temperature for any period of time up to 24 hours prior to addition to the Membrane Device.

3. Using a new transfer pipette, transfer 500 µL of the diluted sample-conjugate mixture into the Sample Well (smaller hole in the top right corner of the device) of a Membrane Device, making certain to expel the liquid sample onto the wicking pad inside of the Membrane Device. When loading the sample into the sample well, make sure that the tip of the transfer pipette is angled towards the Reaction Window (larger hole in the middle of the device).

4. Incubate the device at room temperature for 15 minutes – the sample will wick through the device and a wet area will spread across the Reaction Window.

**NOTE FOR SAMPLES THAT FAIL TO MIGRATE:**
Occasionally, a diluted sample fails to migrate properly and the Reaction Window does not fully wet. If the Reaction Window does not appear to be completely wet within 5 minutes of adding the sample to the Sample Well, then add 100 µL (4 drops) of Diluent to the Sample Well and wait an additional 5 minutes (for a total of 20 minutes).

5. After the incubation, add 300 µL of Wash Buffer to the Reaction Window using the graduated white dropper assembly. Allow the Wash Buffer to flow through the Reaction Window membrane and be absorbed completely.

6. Add 2 drops of Substrate (white-capped bottle) to the Reaction Window. Read and record results visually after 10 minutes.

**INTERPRETATION OF RESULTS**
1. Interpretation of the test is most reliable when the device is read immediately at the end of the 10 minute reaction period. Read the device at a normal working distance in a well-lit area. View with a line of vision directly over the device.

2. Observe device for the appearance of blue dots in the middle of the Reaction Window representing the internal positive control. The appearance of any control dot(s) represents a valid internal control. The background may appear white to light blue in color. Observe device for the appearance of blue lines on the “Ag” and “Tox” sides of the Reaction Window representing the test lines. The lines may appear faint to dark in intensity.

**EFFECT OF FECAL SPECIMEN CONSISTENCY**

**Effect of fecal specimen consistency on the C. DIFF QUIK CHEK COMPLETE® test**

The reaction of fecal specimens of varying consistencies in the antigen portion (n=978) and toxin portion (n=981) of the C. DIFF QUIK CHEK COMPLETE® test is shown in Tables 4 and 5. The percentages of positive reactions using either culture assay or the C. DIFF QUIK CHEK COMPLETE® test were similar in all three types of fecal specimens (liquid, semi-solid, and solid). All of the specimens were submitted for C. difficile testing. The basis of the submission was the clinical history of the patient and not the consistency of the specimen. In the antigen portion, the results show that the C. DIFF QUIK CHEK COMPLETE® test performed similarly to bacterial culture when testing samples of different consistencies. In the toxin portion, the results show the C. DIFF QUIK CHEK COMPLETE® test performed similarly to the tissue culture assay when testing samples of different consistencies.

<table>
<thead>
<tr>
<th>Table 4. Reaction of fecal specimens of varying consistencies in the antigen portion of the C. DIFF QUIK CHEK COMPLETE® test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of specimens</td>
</tr>
<tr>
<td>Positive by bacterial culture assay</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Positive</td>
</tr>
<tr>
<td>Negative by bacterial culture assay</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Antigen Line Negative</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Table 5. Reaction of fecal specimens of varying consistencies in the toxin portion of the C. DIFF QUIK CHEK COMPLETE® test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of specimens</td>
</tr>
<tr>
<td>Positive by tissue culture assay</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Toxin Line Positive</td>
</tr>
<tr>
<td>Negative by tissue culture assay</td>
</tr>
<tr>
<td>C. DIFF QUIK CHEK COMPLETE® Toxin Line Negative</td>
</tr>
</tbody>
</table>
The samples produced the expected results 100% of the time.

An additional 5-day study was performed at 3 sites by running a set of low positive, moderate positive and high negative fecal samples that were spiked with glutamate dehydrogenase, Toxin A and Toxin B. The samples were run in triplicate, twice a day over a 5-day period by multiple technicians at each site. The combined antigen and toxin data from the 5-day reproducibility study is shown in Table 6. The antigen data for Sample 1 was below 90% negative at one site and was consistently negative for antigen at the other two sites. The toxin data for Sample A was below 90% positive at one site, and was consistently positive for toxin at the other two sites. No single site reported results below expectations for both antigen and toxin.

Table 6. Summary of 5-day reproducibility study.

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Antigen Positive</th>
<th>Antigen Negative</th>
<th>Toxin Positive</th>
<th>Toxin Negative</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample 1 (95% negative results expected)</td>
<td>15 (16.7%)</td>
<td>75 (83.3%)</td>
<td>0 (0%)</td>
<td>90 (100%)</td>
</tr>
<tr>
<td>Sample A (95% positive results expected)</td>
<td>89 (98.9%)</td>
<td>1 (1.1%)</td>
<td>65 (72.2%)</td>
<td>25 (27.8%)</td>
</tr>
<tr>
<td>Sample B (mod positive)</td>
<td>87 (96.7%)</td>
<td>3 (3.3%)</td>
<td>86 (95.5%)</td>
<td>4 (4.5%)</td>
</tr>
</tbody>
</table>

CROSS REACTIVITY

Fecal specimens inoculated with the following microorganisms to a final concentration of approximately 10^8 or higher organisms per mL did not react in the antigen or toxin portion of the C. DIFF QUIK CHEK COMPLETE® test:

**Bacterium or Pathogen:** Aeromonas hydrophila, Bacillus cereus, Bacillus subtilis, Bacteroides fragilis, Campylobacter coli, Campylobacter fetus, Campylobacter jejuni, Candida albicans, Clostridium butyricum, Clostridium clostridiforme, Clostridium haemolyticum, Clostridium histolyticum, Clostridium novyi, Clostridium perfringens, Clostridium septicum, Clostridium sordellii (nontoxicogenic), Clostridium sporogenes, Enterobacter aerogenes, Enterobacter cloacae, Enterococcus faecalis, Escherichia coli EIEC, Escherichia coli, Escherichia coli O157:H7, Escherichia coli ETEC, Klebsiella pneumoniae, Proteus vulgaris, Pseudomonas aeruginosa, Salmonella typhimurium, Seratia liquefaciens, Shigella dysenteriae, Shigella flexneri, Shigella sonnei, Staphylococcus aureus, Staphylococcus aureus (Cowan's), Staphylococcus epidermidis, Vibrio cholerae, Vibrio parahaemolyticus, Yersinia enterocolitica

**ANALYTICAL SENSITIVITY**

The cutoff for the assay was established at concentrations of 0.63 ng/mL for toxin A, 0.16 ng/mL for toxin B, and 0.8 ng/mL for glutamate dehydrogenase.

**REPRODUCIBILITY**

The reproducibility of the C. DIFF QUIK CHEK COMPLETE® test was determined using 12 fecal specimens that were coded to prevent their identification during testing. Testing was performed at 3 independent laboratories, which tested the samples for 3 days. The samples produced the expected results 100% of the time.

1. Bring all reagents and the required number of devices to room temperature before use. It is recommended to remove the reagents from the foam insert to reduce the time needed to warm to room temperature.
2. Set up and label one small test tube for each specimen, and optional external controls as necessary.
3. Using the black graduated dropper assembly, add 750 µL (2nd graduation from the tip) Diluent to each tube for fecal specimens.

   **For specimens in transport media such as Cary Blair or C&S, add 650 µL (1st graduation from the tip) of Diluent to the tube.**

4. Add one drop of Conjugate (red capped bottle) to each tube.
5. Obtain one disposable plastic transfer pipette (supplied with the kit) for each sample – the pipettes have raised graduations at 25 µL, 400 µL and 500 µL.

**Graduated Transfer Pipette:**

6. Mix all specimens thoroughly regardless of consistency- it is essential that the specimens be evenly suspended before transferring.

   **Liquid/Semi-solid specimens** – pipette 25 µL of specimen with a transfer pipette and dispense into the Diluent/Conjugate mixture. Use the same transfer pipette to mix the diluted specimen.

   **Formed/Solid specimens** – Care must be taken to add the correct amount of formed feces to the sample mixture. Mix the specimen thoroughly using a wooden applicator stick and transfer a small portion (approximately 2 mm diameter, the equivalent of 25 µL) of the specimen into the Diluent/Conjugate mixture. Emulsify the specimen using the applicator stick.

   **Fecal specimens in Cary Blair or C&S transport media** - pipette 100 µL (2 drops from transfer pipette) of sample into the Diluent/Conjugate mixture.

7. **Optional External Control Samples:**

   **External Positive Control** - add one drop of Positive Control (gray-capped bottle) into the Diluent/Conjugate mixture.

   **External Negative Control** - add 25 µL Diluent into the Diluent/Conjugate mixture.

**NOTE:** Transferring too little specimen, or failure to mix and completely suspend the specimen in the Diluent/Conjugate mixture, may result in a false-negative test result. The addition of too much fecal specimen may cause invalid results due to restricted sample flow.
The test has been optimized for sensitivity and specificity. Alterations of the specified procedure and/or test conditions may affect the sensitivity and specificity of the test. Do not deviate from the specified procedure.

Be attentive to the total assay time when testing more than one fecal specimen. Add Diluent first, and then add the Conjugate to each tube of Diluent. Then add specimen to the tube of Diluent/Conjugate. Thoroughly mix all of the diluted specimens, and transfer to the Membrane Device. The 15-minute incubation step begins after the last diluted sample-conjugate mixture has been transferred to the final Membrane Device.

If the Substrate reagent changes to a dark blue/violet color call technical services for replacement.

Fecal specimens may contain potentially infectious agents and should be handled at “Biosafety Level 2” as recommended in the CDC/NIH Manual “Biosafety in Microbiological and Biomedical Laboratories.”

All reagents are for in vitro diagnostic use only.

Wear disposable gloves when doing the test.

The Diluent reagent contains 0.05% ProClin® 300 as a preservative. Although the concentration is low, ProClin® 300 is known to be harmful. If skin irritation or rash occurs, get medical advice/attention. Take off contaminated clothing and wash it before reuse. Handle reagents according to existing regulations for laboratory safety and good laboratory practice. Safety Data Sheets for this product are available upon request, contact technical support.

Follow your national, regional, and local ordinances accordingly for waste disposal regulations.

### COLLECTION, HANDLING, AND STORAGE OF FECAL SPECIMENS

<table>
<thead>
<tr>
<th>Acceptable Sample Types</th>
<th>Do Not Use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fresh Fecal Specimens</td>
<td>Fecal specimens in Formalin-based fixative (e.g. sodium acetate formalin, 10% formalin, merthiolate formalin)</td>
</tr>
<tr>
<td>Frozen Fecal Specimens</td>
<td>Fecal specimens in alcohol-based fixative (e.g. polyvinyl alcohol)</td>
</tr>
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</table>

<table>
<thead>
<tr>
<th>Sample Storage Temperature</th>
<th>Acceptable length of storage</th>
<th>Comments</th>
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<tr>
<td>2°C – 8°C</td>
<td>72 hours</td>
<td>Ideal specimens are less than 24 hours old</td>
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<td>Longer than 72 hours</td>
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</tr>
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1. Standard collection and handling procedures used in-house for fecal specimens are appropriate.
2. Fecal specimens should be collected in clean, leak-proof containers.
3. Storing fecal specimens in the Diluent is NOT recommended.
4. Do not allow the fecal specimens to remain in the Diluent/Conjugate mixture for >24 hours.

The only non-\textit{C. difficile} organism to react in the toxin portion of the \textit{C. DIFF QUIK CHEK COMPLETE}® test was \textit{Clostridium sordellii} VPI 9048. This strain produces toxins HT and LT, which are homologous to toxins A and B, respectively.

The following viruses of $10^{3.3}$ to $10^{8.25}$ TCID units per 0.2 mL did not react in the \textit{C. DIFF QUIK CHEK COMPLETE}® test:

**Viruses:** Adenovirus types 1, 2, 3, 5, 40, 41, Human coronavirus, Coxackievirus B2, B3, B4, B5, Echovirus 9, 11, 18, 22, 33, Enterovirus type 68, 69, 70, 71, Rotavirus.

### INTERFERING SUBSTANCES

The following substances (U.S. formulation) had no effect on test results when present in feces in the concentrations indicated: mucin (3.5% w/v), human blood (40% v/v), barium sulfate (5% w/v), Imodium® (5% v/v), Kapectate® (5% v/v), Pepto-Bismol® (5% v/v), steric/palmitic acid (40% w/v), Metronidazole (0.25% w/v), Vancomycin (0.25% w/v).

### REACTION OF CLINICAL ISOLATES OBTAINED ON CYCLOSERINE-CEFOXITIN-FRUCTOSE AGAR (CCFA)

A total of 103 \textit{C. difficile} clinical isolates, obtained by anaerobic bacterial culture on CCFA after 3 days at 37°C, were tested in the \textit{C. DIFF QUIK CHEK COMPLETE}® test. For the analysis, individual colonies were picked and suspended in Diluent as recommended for fecal specimens. All 103 isolates gave a positive antigen reaction in the test.

Seventy of the 103 isolates (68%) were from fecal specimens that were positive for \textit{C. difficile} toxin by tissue culture assay. Of these, 56 (80%) gave a positive toxin reaction when screened following anaerobic growth on CCFA for 3 days at 37°C.

1. 4
2. 13
3. 11.
4. The test has been optimized for sensitivity and specificity. Alterations of the specified procedure and/or test conditions may affect the sensitivity and specificity of the test. Do not deviate from the specified procedure.
5. Be attentive to the total assay time when testing more than one fecal specimen. Add Diluent first, and then add the Conjugate to each tube of Diluent. Then add specimen to the tube of Diluent/Conjugate. Thoroughly mix all of the diluted specimens, and transfer to the Membrane Device. The 15-minute incubation step begins after the last diluted sample-conjugate mixture has been transferred to the final Membrane Device.
6. If the Substrate reagent changes to a dark blue/violet color call technical services for replacement.
7. Fecal specimens may contain potentially infectious agents and should be handled at “Biosafety Level 2” as recommended in the CDC/NIH Manual “Biosafety in Microbiological and Biomedical Laboratories.”
8. All reagents are for in vitro diagnostic use only.
9. Wear disposable gloves when doing the test.
10. The Diluent reagent contains 0.05% ProClin® 300 as a preservative. Although the concentration is low, ProClin® 300 is known to be harmful. If skin irritation or rash occurs, get medical advice/attention. Take off contaminated clothing and wash it before reuse. Handle reagents according to existing regulations for laboratory safety and good laboratory practice. Safety Data Sheets for this product are available upon request, contact technical support.
11. Follow your national, regional, and local ordinances accordingly for waste disposal regulations.

### COLLECTION, HANDLING, AND STORAGE OF FECAL SPECIMENS

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REFERENCES

MATERIALS PROVIDED

Membrane Devices – each pouch contains 1 device
Diluent (22 mL per bottle) – Buffed protein solution with graduated dropper assembly (contains 0.05% ProClin® 300)
Signal Word: Warning
H317: May cause an allergic skin reaction
P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501

Wash Buffer (12 mL per bottle) – Buffered solution with graduated dropper assembly (contains 0.05% ProClin® 300)
Signal Word: Warning
H317: May cause an allergic skin reaction
P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501

Substrate (3.5 mL per bottle) – Solution containing tetramethylbenzidine Conjugate (2.5 mL per bottle) – Mouse monoclonal antibody specific for glutamate dehydrogenase coupled to horseradish peroxidase and goat polyclonal antibodies specific for toxins A and B coupled to horseradish peroxidase in a buffered protein solution (contains 0.05% ProClin® 300)
Signal Word: Warning
H317: May cause an allergic skin reaction
P261, P272, P280, P302, P352, P333, P313, P321, P362, P364, P501

Positive Control (2 mL) – Antigen in a buffered protein solution
Disposable plastic transfer pipettes – graduated at 25 µL, 400 µL and 500 µL

In Vitro Diagnostic Medical Device

MATERIALS AND EQUIPMENT REQUIRED BUT NOT PROVIDED
Small test tubes (e.g., plastic Eppendorf tubes or glass tubes)
Applicator sticks
Timer
Disposable gloves for handling fecal samples
Vortex mixer
Pipettor and tips

SHELF LIFE AND STORAGE
The expiration date of the kit is given on the label. Expiration dates for each component are listed on the individual labels. The kit should be stored between 2°C and 8°C.

PRECAUTIONS
1. Reagents from different kits should not be mixed or interchanged. Do not use a kit past the expiration date.
2. Each component in the kit should be inspected for any signs of leakage. Upon arrival, inspect the kit to ensure that components are not frozen or warm to the touch due to improper shipping conditions.
3. Bring all components to ROOM TEMPERATURE BEFORE USE!
4. Caps, tips and dropper assemblies are color-coded; do NOT mix or interchange!
5. Do not freeze the reagents. The kit should be stored between 2°C and 8°C.
6. The pouch containing the Membrane Device should be at room temperature before opening. Keep the membrane devices dry before use.
7. Use fecal specimens within 72 hours of collection to obtain optimal results. Specimens that are frozen may lose activity due to freezing and thawing. If using frozen specimens, thaw at room temperature.
8. Hold reagent bottles vertically to dispense reagents to ensure consistent drop size and correct volume.
9. Specimens and membrane devices should be handled and disposed of as potential biohazards after use.
10. Membrane devices cannot be reused.
INTENDED USE

The C. DIFF QUIK CHEK COMPLETE® test is a rapid membrane enzyme immunoassay for the simultaneous detection of *Clostridium difficile* glutamate dehydrogenase antigen and toxins A and B in a single reaction well. The test detects *C. difficile* antigen, glutamate dehydrogenase, as a screen for the presence of *C. difficile* and confirms the presence of toxigenic *C. difficile* by detecting toxins A and B in fecal specimens from persons suspected of having *C. difficile* disease. The test is to be used as an aid in the diagnosis of *C. difficile* disease. As with other *C. difficile* tests, results should be considered in conjunction with the patient history.

EXPLANATION

After treatment with antibiotics, many patients develop gastrointestinal problems ranging from mild diarrhea to severe pseudomembranous colitis. Many cases of the milder forms of gastrointestinal illness and most cases of pseudomembranous colitis are caused by toxigenic strains of *Clostridium difficile* (1). This organism is an opportunistic anaerobic bacterium that grows in the intestine once the normal flora has been altered by the antibiotic. Toxigenic strains of *C. difficile* carry the genes encoding the toxins while non-toxigenic strains do not carry the toxin genes. Disease onset is associated with the toxins that are produced by the toxigenic organism. The clinical symptoms associated with the disease are believed to be primarily due to toxin A, which is a tissue-damaging enterotoxin (2,3). *C. difficile* also produces a second toxin, designated toxin B. Toxin B, which has been referred to as the cytotoxin of the organism, is the toxin detected by the tissue culture assay currently used by many laboratories. Toxigenic *C. difficile* strains produce both toxins, or only toxin B (4-7). The glutamate dehydrogenase of *C. difficile* is a good antigen marker for the organism in feces because it is produced in high amounts by all strains, toxigenic or non-toxigenic (8-10). The antigen can be detected in fecal specimens by using the C. DIFF QUIK CHEK COMPLETE® test. A positive result in the test for the glutamate dehydrogenase of *C. difficile* confirms the presence of this organism in a fecal specimen; a negative result indicates the absence of the organism. A positive result in the test for toxins A and B confirms the presence of toxigenic *C. difficile*.

PRINCIPLE OF THE TEST

The C. DIFF QUIK CHEK COMPLETE® test uses antibodies specific for glutamate dehydrogenase and toxins A and B of *C. difficile*. The device contains a Reaction Window with three vertical lines of immobilized antibodies. The antigen test line ("Ag") contains antibodies against *C. difficile* glutamate dehydrogenase. The control line ("C") is a dotted line that contains anti-horseradish peroxidase (HRP) antibodies. The toxins A and B test line ("Tox") contains antibodies against *C. difficile* toxins A and B. The Conjugate consists of antibodies to glutamate dehydrogenase and antibodies to toxins A and B coupled to horseradish peroxidase. To perform the test, the sample is added to a tube containing a mixture of Diluent and Conjugate. The diluted sample-conjugate mixture is added to the Sample Well and the device is allowed to incubate at room temperature for 15 minutes. During the incubation, any glutamate dehydrogenase and toxins A and B in the sample bind to the antibody-peroxidase conjugates. The antibody-antibody-conjugate complexes migrate through a filter pad to a membrane where they are captured by the immobilized glutamate dehydrogenase-specific and toxins A and B-specific antibodies in the lines. The Reaction Window is subsequently washed with Wash Buffer, followed by the addition of Substrate. After a 10 minute incubation period, the "Ag" reaction is examined visually for the appearance of a vertical blue line on the "Ag" side of the Reaction Window. A blue line indicates a positive test. If the "Ag" is positive, then the "Tox" reaction should be examined visually for the appearance of a blue line on the "Tox" side of the Reaction Window. A blue line indicates a positive test. A positive "C" reaction, indicated by a vertical dotted blue line under the "C" portion of the Reaction Window, confirms that the test is working properly and the results are valid.
A Rapid Membrane Enzyme Immunoassay for the Simultaneous Detection of Clostridium difficile Glutamate Dehydrogenase Antigen and Toxins A and B in Fecal Specimens

Catalog No. 30525C (25 Tests) or 30550C (50 Tests)

In Vitro Diagnostic Medical Device

U.S. CLIA classification — Moderate

U. S. Patent #8,343,726